

Position paper.

## Ergot (*Claviceps purpurea*) – An aposematic fungus

Simcha Lev-Yadun\* and Malka Halpern

Department of Biology, Faculty of Science and Science Education, University of Haifa - Oranim, Tivon 36006, Israel, Tel. +972-4-9838827, Fax +972-4-9832167, Email. lev Yadun@research.haifa.ac.il and halpern@macam.ac.il

(Received December 18, 2006; Accepted March 18, 2007)

### Abstract

Predators learn to associate conspicuous signals, such as bright colors, with unpalatability or danger. This defensive strategy is termed Aposematism. We propose that the very poisonous purple-black sclerotia of the infamous fungus *Claviceps purpurea* (ergot) and many other *Claviceps* species are aposematic. Very toxic fungal sclerotia are associated with conspicuous colors (black, yellow, purple, reddish, brown, violet, white and their combinations) and severely harm herbivores that consume the infected plants, thus meeting the criteria for aposematism. Fungi, that only moderately reduce the reproductive capacity of their hosts can protect the host plants from herbivory and weaken the evolutionary tendency of their hosts to evolve better resistance to infection.

**Keywords:** Aposematic coloration, *Claviceps*, defense, ergot, herbivory, mutualism

### 1. Introduction

Aposematic coloration is a well known and widespread defensive phenomenon found across the animal kingdom and in many vascular plants. Aposematism is the phenomenon evolved by organisms that are unpalatable or dangerous to potential predators/herbivores, whereby they advertise their bad quality as a food source by a variety of colors, mainly bright and/or contrasting, e.g., red, orange, yellow, black, white, and their combinations (e.g., Cott, 1940; Edmunds, 1974; Gittleman and Harvey, 1980; Lev-Yadun, 2001, 2006; Lev-Yadun and Ne'eman, 2004, 2006; Ruxton et al., 2004) or by odor (Eisner and Grant, 1981). This facilitates associative learning that reduces further use. The vast majority of studies on animal aposematism associate warning coloration with chemical defense (e.g., Edmunds, 1974; Gittleman and Harvey, 1980; Harvey and Paxton, 1981; Ruxton et al., 2004). The common aposematism of spiny animals received much less attention (Cott, 1940; Ruxton et al., 2004; Inbar and Lev-Yadun, 2005; Speed and Ruxton, 2005). In plants, most attention with respect to aposematic coloration has focused on its association with mechanical

defense by thorns (Lev-Yadun, 2001, 2003a,b, 2006; Lev-Yadun and Ne'eman, 2004, 2006; Rubino and McCarthy, 2004; Ruxton et al., 2004; Halpern et al., 2007) although certain poisonous plants have been considered as aposematic (Cook et al., 1971; Hinton, 1973; Wiens, 1978; Harborne, 1982; Lev-Yadun and Ne'eman, 2004; Lev-Yadun, 2006). That mushrooms are also aposematic has been hypothesized, however, while some poisonous mushrooms are colorful, many edible ones are similarly colorful. For this reason, the idea of aposematic coloration in fungi (Camazine, 1983) has not been supported as a general phenomenon by field data and taxonomic analysis (Guevara and Dirzo, 1999; Sherratt et al., 2005). Taste and odor seem to be more important as a common aposematic signals in large fungi, probably since many of the animals that usually consume them are nocturnal and because low level of illumination on the forest floor would reduce the effectiveness of bright coloration as a warning signal (Camazine, 1983, 1985; Guevara and Dirzo, 1999; Sherratt et al., 2005).

### 2. Ergot

Ergot, a fungal disease of rye and other cereals, caused by

\*The author to whom correspondence should be sent.

the fungus *Claviceps purpurea*, is one of the most notorious fungi in human history (Matossian, 1989; Bennett and Bentley, 1999; Alm, 2003). There are more than 40 species in the genus *Claviceps* (Pažoutová and Parbery, 1999) which attack grasses, rushes, and sedges. Most ergot species have colorful and conspicuous sclerotia that form on the ears of their host (Tenberge, 1999). Ergot sclerotia develop in only some of the flowers of their host, allowing the infected plants to produce a considerable number of viable seeds. The fungal sclerotia, are rich in a wide array of highly poisonous alkaloids (Mantle, 1969; Lorenz, 1979; Matossian, 1989; van Dongen and de Groot, 1995). Ergotism, the toxic condition in humans and animals that eat *Claviceps*-infected grain, is often accompanied by psychotic delusions, nervous spasms, abortion, convulsions, gangrene, and even death (Mantle, 1969; Lorenz, 1979; Matossian, 1989; van Dongen and de Groot, 1995). The best-known species, *C. purpurea*, forms conspicuous purple-black sclerotia up to several-cm-long on the ears of rye, where they are easily seen. When infected rye (a staple for humans in European countries with cold wet climates) was ground and used to produce bread, non-lethal levels of ergot poisoning caused severe hallucinations or intense burning pain (St Anthony's Fire) and gangrene of feet, hands, and whole limbs, due to the vasoconstrictive action of the ergot alkaloids (De Costa, 2002). Historically, ergotism has killed many thousands of people. For instance, over 40,000 deaths attributed to ergot poisoning were recorded in France in 943 (Prescott et al., 2005). Many witch-trials, and subsequent executions, in medieval Europe over centuries, and possibly some in North America involved women who did not behave according to acceptable norms, most likely as a result of intoxication by ergot alkaloids (Matossian, 1989; Alm, 2003). The pharmacological activities of the fungus are due to components that include lysergic acid diethylamide (LSD) (Matossian, 1989; Bennett and Bentley, 1999; Alm, 2003; Eadie, 2004). People learned to refrain from eating infected rye, and it is likely that herbivores that consume grasses would learn to avoid it too. The antiherbivory functions of ergot alkaloids, against both invertebrates and vertebrates, have led to it being regarded as mutualism: the plants are defended from herbivory by the poisonous substances of the fungi, while the fungi receive nutrition from the host (Clay, 1988).

### 3. Discussion

We propose that the very poisonous and colorful sclerotia of *Claviceps* species are aposematic. Sclerotia of *Claviceps hirtella* are yellow, those of *C. glabra* brown, those of *C. viridis* green, and those of *C. purpurea* dark purple or black (Lorenz, 1979) – typical colors of poisonous aposematic organisms. Pažoutová (2006) provides detailed data on the

colors of sclerotia for most *Claviceps* species on this website. Eighteen species have black or blackish sclerotia, in eight the sclerotia are of various shades of brown, in seven they are yellow, four have red or purple sclerotia, one has green, and one is partly white. Thus, the majority of the *Claviceps* species have conspicuous sclerotia which are also poisonous. The association of unpalatable, very toxic fungal organs (sclerotia) with conspicuous colors (black, yellow, purple, reddish, brown, violet, white, and their combinations), and the well-documented toxicity to herbivores that consume them, meet the criteria for characterizing the association between *Claviceps* and grasses as operative aposematism. Poisonous mollusks, insects or reptiles with bright colors are considered aposematic without experimental data concerning deterrence of their predators. The same should apply when fungi are discussed.

As for the question do large herbivores (sheep, goat, deer, cattle, etc.) pay attention to patterns of plant coloration at the size of ergot sclerotia, the answer is yes. A field experiment, by Cahn and Harper (1976), showed clearly that rumen-fistulated sheep, which could be directly sampled for diet-content, preferred unmarked leaves of *Trifolium repens* over marked (variegated) ones, indicating the probable defensive value of such variegation. Ergot infests cereals which grow in open habitats where grazers usually feed during daytime, optimal conditions for visual aposematism. The function of bright fungal colors deserves more attention. In plants for instance, various pigments may simultaneously serve defense, physiological functions and attraction of pollinators or seed dispersers (Gould, 2004; Lev-Yadun et al., 2002, 2004; Schaefer and Wilkinson, 2004; Lev-Yadun, 2006) and fungal coloration may similarly have more than one function.

The plant/*Claviceps* interactions may be viewed simply as a disease, or as a mutualistic relationship. Defensive mutualism between plants and fungi is well known (Clay, 1988; Bush et al., 1997; Omacinl et al., 2001; Clay and Scharld, 2002), but has not been discussed as an aposematic effect. Fungal endophytes in the genus *Neotyphodium* (Ascomycetes: Clavicipitaceae) form mutualistic associations with a variety of grasses (Clay, 1990; Breen, 1994). The fungal hyphae grow intercellularly in leaf and stem tissues, causing infections that are transmitted exclusively through the seeds of the host plant. The fungus benefits from access to plant nutrient and photosynthetic resources, while the plant benefits from enhanced resistance to insect herbivores or vertebrate grazers (Clay, 1990; Breen, 1994; Elmi and West, 1995). A series of fungal endophyte-mediated-alkaloids provides the basis for the acquired chemical defense against herbivory (Porter, 1994; Justus et al., 1997). Ergot fungi parasitize rye and other grasses, reducing their reproduction, the very poisonous fungus harming the herbivores that eat the infected plants. Potentially, this benefits individual plants, and probably nearby plants too.

The overall gain to the host seems to more than compensate for the moderate reduction in its reproduction.

Since *Claviceps* species need plant hosts, the more toxic the ergots are to herbivores, the better for both ergot and host. Chemicals in the sclerotia may also directly prevent their consumption by the herbivores or from attacks by other fungi and microorganisms. Indeed, large herbivores usually learn to avoid toxic plants (e.g., Howe and Westley, 1988; Lev-Yadun and Ne'eman, 2004), resulting in a reduced tendency to consume infected hosts. For instance, Opossums (*Didelphis virginiana*), are known to develop an aversion towards the very poisonous mushroom *Amanita muscaria*, in part due to the toxin muscimol (Camazine, 1983). Alternatively, ergot fungi may induce abortions or kill the animals that eat ergot-infested species, decreasing grazing pressure. Since low levels of ergot consumption are not lethal, but cause sickness (Clay, 1988; Matossian, 1989), conditions are appropriate for the development of food aversion towards ergot-infected grasses. The array of alkaloids in ergots makes it difficult for herbivores to evolve resistance to ergot toxicity. When ergot fungi protect their host populations against herbivory, they protect their own habitat and benefiting the host plants. They also reduce the host's evolutionary tendency to enhance its resistance to the disease. In our view the association between *Claviceps* and grasses is a fine-tuned ecological tactic that fits the definition of "dangerous liaisons" sensu van Baalen and Jansen (2001). We propose that odor, in addition to color, might be involved in the aposematic signaling of ergots even in their open, well-illuminated habitats as was found for fungi that grow in dark habitats as described by Camazine (1983, 1985), Guevara and Dirzo (1999) and Sherratt et al. (2005).

The hypothesis we present here may apply only to the fungus *Claviceps* that form ergots, or have a broader scope. There are indications for the latter since other fungal taxa express potential aposematic coloration. For example, *Aspergillus* species produce aflatoxin, a fungal metabolite which is a very potent toxin if consumed by animals (Payne and Brown, 1998) and have toxic pigmented sclerotia (black in *A. flavus* and *A. tamarii*) (Raper and Fennell, 1965; Goto et al., 1996; Chang et al., 2001). A similar picture emerges in the genus *Fusarium* (e.g., Toussoun and Nelson, 1968; Bottalico, 1998), as various animals are repulsed by food contaminated with its toxins (Mirocha et al., 1976). Finally, the insect pathogenic fungus *Cordyceps* produces several types of pigments some of which are bright red (Unagul et al., 2005) and seems to fit into the general picture of toxicity associated with bright or contrasting coloration. The lack of an explanation for the association of toxicity with coloration in fungi, is just one reason why these fungi are worthy of further study. We hope that our hypothesis will stimulate observational and experimental research on this overlooked aspect in fungal biology.

## Acknowledgements

We thank David Richardson and an anonymous reviewer for their helpful comments on this manuscript.

## REFERENCES

- Alm, T. 2003. The witch trials of Finnmark, northern Norway, during the 17th century: Evidence for ergotism as a contributing factor. *Economic Botany* **57**: 403–416.
- Bennett, J.W. and Bentley, R. 1999. Pride and prejudice: the story of ergot. *Perspectives in Biology and Medicine* **42**: 333–355.
- Bottalico, A. 1998. *Fusarium* diseases of cereals: Species complex and related mycotoxin profiles, in Europe. *Journal of Plant Pathology* **80**: 85–103.
- Breen, J.P. 1994. *Acremonium* endophyte interactions with enhanced plant resistance to insects. *Annual Review of Entomology* **39**: 401–423.
- Bush, L.P., Wilkinson, H.H., and Schardl, C.L. 1997. Bioprotective alkaloids of grass-fungal endophyte symbioses. *Plant Physiology* **114**: 1–7.
- Cahn, M.G. and Harper, J.L. 1976. The biology of the leaf mark polymorphism in *Trifolium repens* L. 2. Evidence for the selection of leaf marks by rumen fistulated sheep. *Heredity* **37**: 327–333.
- Camazine, S. 1983. Mushroom chemical defense: food aversion learning induced by hallucinogenic toxin, muscimol. *Journal of Chemical Ecology* **9**: 1473–1481.
- Camazine, S. 1985. Olfactory aposematism: association of food toxicity with naturally occurring odor. *Journal of Chemical Ecology* **11**: 1289–1295.
- Chang, P.-K., Bennett, J.W., and Cotty, P.J. 2001. Association of aflatoxin biosynthesis and sclerotial development in *Aspergillus parasiticus*. *Mycopathologia* **153**: 41–48.
- Clay, K. 1988. Clavicipitaceous fungal endophytes of grasses: coevolution and the change from parasitism to mutualism. In: *Coevolution of Fungi with Plants and Animals*. Pirozynski, K.A., and Hawksworth, D.L., eds. Academic Press, London, pp. 79–105.
- Clay, K. 1990. Fungal endophytes of grasses. *Annual Review of Ecology and Systematics* **21**: 275–297.
- Clay, K. and Schardl, C. 2002. Evolutionary origins and ecological consequences of endophyte symbiosis with grasses. *American Naturalist* **160** (Supplement): S99–S127.
- Cook, A.D., Atsatt, P.R., and Simon, C.A. 1971. Doves and dove weed: multiple defenses against avian predation. *BioScience* **21**: 277–281.
- Cott, H.B. 1940. *Adaptive Coloration in Animals*. Methuen & Co. Ltd., London, pp. 508 + 48 plates.
- De Costa, C. 2002. St Anthony's fire and living ligatures: A short history of ergometrine. *Lancet* **359**: 1768–1770.
- Eadie, M.J. 2004. Ergot of rye – The first specific for migraine. *Journal of Clinical Neuroscience* **11**: 4–7.
- Edmunds, M. 1974. *Defence in Animals. A survey of anti-predator defences*. Longman Group Ltd., Harlow, pp. 357.
- Eisner, T. and Grant, R.P. 1981. Toxicity, odor aversion, and "olfactory aposematism". *Science* **213**: 476.
- Elmi, A.A. and West, C.P. 1995. Endophyte infection effects on stomatal conductance, osmotic adjustment, and drought recovery of tall fescue. *New Phytologist* **131**: 61–67.
- Gittleman, J.L. and Harvey, P.H. 1980. Why are distasteful prey not cryptic? *Nature* **286**: 149–150.

- Givnish, T.J. 1990. Leaf mottling: relation to growth form and leaf phenology and possible role as camouflage. *Functional Ecology* **4**: 463–474.
- Goto, T., Wicklow, D.T., and Ito, Y. 1996. Aflatoxin and cyclopiazonic acid production by a sclerotium-producing *Aspergillus tamarii* strain. *Applied and Environmental Microbiology* **62**: 4036–4038.
- Gould, K.S. 2004. Nature's Swiss army knife: The diverse protective roles of anthocyanins in leaves. *Journal of Biomedicine & Biotechnology* **2004**: 314–320.
- Guevara, R. and Dirzo, R. 1999. Consumption of macro-fungi by invertebrates in a Mexican tropical cloud forest: do fruit body characteristics matter? *Journal of Tropical Ecology* **15**: 603–617.
- Halpern, M., Raats, D., and Lev-Yadun, S. 2007. Plant biological warfare: Thorns inject pathogenic bacteria into herbivores. *Environmental Microbiology* **9**: 584–592.
- Harborne, J.B. 1982. *Introduction to Ecological Biochemistry*. 2nd ed. Academic Press, London, pp. 278.
- Harvey, P.H. and Paxton, R.J. 1981. The evolution of aposematic coloration. *Oikos* **37**: 391–396.
- Hinton, H.E. 1973. Natural deception. In: *Illusion in Nature and Art*. Gregory, R.L. and Gombrich, E.H., eds. Duckworth, London, pp. 97–159.
- Howe, H.F. and Westley, L.C. 1988. *Ecological Relationships of Plants and Animals*. Oxford University Press, New York, pp. 273.
- Inbar, M. and Lev-Yadun, S. 2005. Conspicuous and aposematic spines in the animal kingdom. *Naturwissenschaften* **92**: 170–172.
- Justus, M., Witte, L., and Hartmann, T. 1997. Levels and tissue distribution of loline alkaloids in endophyte-infected *Festuca pratensis*. *Phytochemistry* **44**: 51–57.
- Lev-Yadun, S. 2001. Aposematic (warning) coloration associated with thorns in higher plants. *Journal of Theoretical Biology* **210**: 385–388.
- Lev-Yadun, S. 2003a. Why do some thorny plants resemble green zebras? *Journal of Theoretical Biology* **244**: 483–489.
- Lev-Yadun, S. 2003b. Weapon (thorn) automimicry and mimicry of aposematic colorful thorns in plants. *Journal of Theoretical Biology* **244**: 183–188.
- Lev-Yadun, S. 2006. Defensive coloration in plants: a review of current ideas about anti-herbivore coloration strategies. In: *Floriculture, Ornamental and Plant Biotechnology: Advances and Topical Issues*. Teixeira da Silva, J.A., ed. Global Science Books, London, pp. 292–299.
- Lev-Yadun, S. and Ne'eman, G. 2004. When may green plants be aposematic? *Biological Journal of the Linnean Society* **81**: 413–416.
- Lev-Yadun, S. and Ne'eman, G. 2006. Color changes in old aposematic thorns, spines, and prickles. *Israel Journal of Plant Sciences* (in press).
- Lev-Yadun, S., Dafni, A., Flaishman, M.A., Inbar, M., Izhaki, I., Katzir, G., and Ne'eman, G. 2004. Plant coloration undermines herbivorous insect camouflage. *BioEssays* **26**: 1126–1130.
- Lev-Yadun, S., Dafni, A., Inbar, M., Izhaki, I., and Ne'eman, G. 2002. Colour patterns in vegetative parts of plants deserve more research attention. *Trends in Plant Science* **7**: 59–60.
- Lorenz, K. 1979. Ergot on cereal grains. *Critical Reviews of Food Science and Nutrition* **11**: 311–354.
- Mantle, P.G. 1969. The role of alkaloids in the poisoning of mammals by sclerotia of *Claviceps* spp. *Journal of Stored Products Research* **5**: 237–244.
- Matossian, M.K. 1989. *Poisons of the Past. Molds, Epidemics, and History*. Yale University Press, New Haven, pp. 190.
- Mirocha, C.J., Pathre, S.V., Schauerhamer, B., and Christensen, C.M. 1976. Natural occurrence of *Fusarium* toxins in feedstuff. *Applied and Environmental Microbiology* **32**: 553–556.
- Omacinl, M., Chaneton, E.J., Ghersa, C.M., and Müller, C.B. 2001. Symbiotic fungal endophytes control insect host-parasite interaction webs. *Nature* **409**: 78–81.
- Payne, G.A. and Brown, M.P. 1998. Genetics and physiology of aflatoxin biosynthesis. *Annual Review of Phytopathology* **36**: 329–362.
- Pažoutová, S. 2006. Website at the Institute of Microbiology, Academy of Sciences of the Czech Republic ("Ergot infopage, what is *Claviceps*", the table).
- Pažoutová, S., and Parbery, D.P. 1999. The taxonomy and phylogeny of *Claviceps*. In: *Ergot the Genus Claviceps*. Křen, V. and Cvak, L., eds. Harwood Academic Publishers, Australia, pp. 57–77.
- Porter, J.K. 1994. Chemical constituents of grass endophytes. In: *Biotechnology of Endophytic Fungi of Grasses*. Bacon, C.W. and White, J.F., eds. CRC Press, Boca Raton, pp. 103–123.
- Prescott, L.M., Harley, J.P., and Klein, D.A. 2005. The fungi (Eumycota), slime molds and water molds. In: *Microbiology*. 5th ed. Prescott, L.M., Harley, J.P., and Klein, D.A., eds. McGraw-Hill, New York, pp. 537–552.
- Raper, K.B. and Fennell, D.I. 1965. *The Genus Aspergillus*. Robert E. Krieger Publishing Company, Huntington, 686 pp.
- Rubino, D.L. and McCarthy, B.C. 2004. Presence of aposematic (warning) coloration in vascular plants of southeastern Ohio. *Journal of the Torrey Botanical Society* **131**: 252–256.
- Ruxton, G.D., Sherratt, T.N., and Speed, M.P. 2004. *Avoiding Attack. The Evolutionary Ecology of Crypsis, Warning Signals & Mimicry*. Oxford University Press, Oxford, 249 pp.
- Schaefer, H.M. and Wilkinson, D.M. 2004. Red leaves, insects and coevolution: a red herring? *Trends in Ecology and Evolution* **19**: 616–618.
- Sherratt, T.N., Wilkinson, D.M., and Bain, R.S. 2005. Explaining Dioscorides' "double difference": Why are some mushrooms poisonous, and do they signal their unprofitability? *American Naturalist* **166**: 767–775.
- Speed, M.P. and Ruxton, G.D. 2005. Warning displays in spiny animals: one (more) evolutionary route to aposematism. *Evolution* **59**: 2499–2508.
- Tenberge, K.B. 1999. Biology and life strategy of the ergot fungi. In: *Ergot the Genus Claviceps*. Křen, V. and Cvak, L., eds. Harwood Academic Publishers, Australia, pp. 25–56.
- Toussoun, T.A. and Nelson, P.E. 1968. *A Pictorial Guide to the Identification of Fusarium Species according to the Taxonomic System of Snyder and Hansen*. The Pennsylvania State University Press, University Park, PA, 51 pp.
- Unagul, P., Wongsu, P., Kittakoop, P., Intamas, S., Srikitikulchai, P., and Tanticharoen, M. 2005. Production of red pigments by the insect pathogenic fungus *Cordyceps unilateralis* BCC 1869. *Journal Industrial Microbiology and Biotechnology* **32**: 135–140.
- van Baalen, M. and Jansen, V.A.A. 2001. Dangerous liaisons: The ecology of private interest and common good. *Oikos* **95**: 211–224.
- van Dongen, P.W.J. and de Groot, A.N.J.A. 1995. History of ergot alkaloids from ergotism to ergometrine. *European Journal of Obstetrics & Gynecology and Reproductive Biology* **60**: 109–116.
- Wiens, D. 1978. Mimicry in plants. *Evolutionary Biology* **11**: 365–403.